

The Molecular Basis of their Electrical Excitability

Viva La Complexity!

Consider,

- The human brain contains $>10^{11}$ neurons!
- Each neuron makes 10³ (average) synaptic contacts on up to 10³ other neurons, and receives up to 10⁵ (typically 10⁴) synaptic inputs!
- **Conclusion:** the computational power of the nervous system is based on *sheer numbers of, and connections between, individual elements, as well as in the computational powers of the elements.* Adding modulation and plasticity to each of these connections makes it even more impressive.

Support Staff

CNS: brain and spinal cordPNS: nerves (what are they?)neurons:glia# 1:9volume 1:1

In central nervous system (CNS)

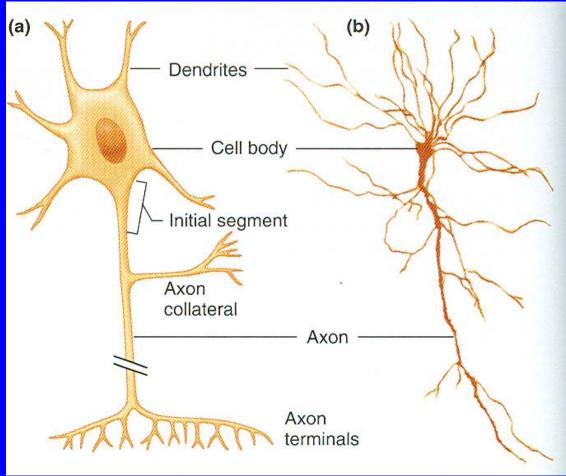
oligodendrocytes: myelinate (up to 40 different) axons *astroglia:* regulates extracellular fluid (K⁺, neurotransmitters, glucose, NH₃, guidance) *microglia:* immune-like macrophagish cells

In peripheral nervous system (*PNS*) *Schwann cells*: myelinate single axons *nodes of Ranvier:* bare axons between myelination

Axons may be 1 m long! nutrients? axonal transport via microtubular "railways", consuming ATP

Neurons Are Cells

- cell body
- organelles
 - nucleus
 - cytoplasm
 - mitochondria
 - endoplasmic reticulum
 - golgi apparatus
- cell processes
 - axon
 - dendrites
- cell membrane: phospholipid bilayer



Neuron Parts by Function

- "Input": chemoelectrical reception; in dendrites (soma, axon)
- "Metabolic Center" soma (axons, dendrites):
 nucleus
 - protein synthesis
 - ATP production
- "Trigger" axon hillock: digital output of spatiotemporal integration
- "Transmission" axon, terminals: transmit information to other cells via action potentials

Distinctive Feature

- In addition to all the genetic and biochemical complexities that all cells possess, neurons have the added feature of having *electrically excitable membranes*, a trait they share with certain other cell types (e.g., myocytes).
- Electrical excitability results from maintenance of membrane potential and differential distribution of many voltage- and ligand-sensitive ion channels in the membrane.
- Communication between cells requires electrical responsivity

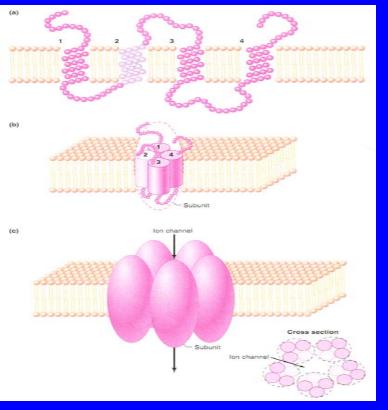
Membrane Function

- physical barrier: holds in contents
- chemical barrier: allows for selective passage of compounds, ions, water.
- amphipathic (polar & non-polar regions)
- composition:
 - phospholipids
 - cholesterol
 - integral membrane proteins
- capacitor: 1 μF/cm²
- membrane potential, typically ~ -65 mV, ~7nm
- Integral membrane proteins: receptors; ion channels;

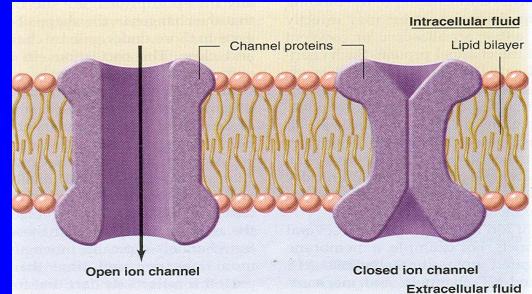
Ion Channels

- Na⁺, K⁺, Cl⁻, Ca²⁺ diffuse across membranes at high rates, despite their ionic nature. How?
- Integral membrane proteins form channels through which ions may pass through membranes
- May be single proteins or protein complexes
- Different channels are selective for different ions
- Pore is v. small: ions in single file

Ion Channels



consist of single proteins or protein complexes



open or close via conformational changes

Ion Flux: Electrical Forces

- electrical potential exists across cell membrane (inside negative relative to outside)
- ions are charged, and thus are influenced by electric field
- consider, if no Δ [in] vs. [out], still have electrical driving force
 - cations into the cell, anions out of the cell
- *electrochemical* gradient takes into account concentration gradient *and* electrical potential
- forces may act in concert *or* oppose each other

A little more complex...

- Ion channels may be open or closed, based on conformational change of the protein
- Process may be very rapid: $\tau < 0.5$ ms
- At a given EC gradient and time period, total ion flow depends on the frequency and duration of it being open (i.e., it's probability of being open) x driving force
- Probability depends on:
 - allosteric/covalent forces for *ligand-gated* channels
 - membrane potential in *voltage-gated* channels
 - physical deformation in *mechanically-gated* channels
 - a combination of the above

Biophysics: Resting E_m

- cells maintain negative intracellular potential via ion pumping
- membrane has conductive ion channels
- membrane also acts as capacitor
- ion separation = electrochemical gradient:

 $E = [RT/(z_sF)]ln([out]/[in]) Nernst, 1888$

R, gas constant; F, faraday; T, temp; z_s, valence

 $E \sim 61.54\log([out]/[in])$ (@body temp, 1 atm)

Ion	[in] (mM)	[out](mM)	E _{Nernst} (mV)	EG (-60mV)	CG	Net
Na ⁺	15	150	+62	+122 in	in	in
K ⁺	150	5.5	-89	-29 in	out	out
Cŀ	9	125	-71	-11 out	in	0
Ca ²⁺	.0001	1.5	+129	+189 in	in	in

must then multiply Net by conductance to get flow

Goldman Equation

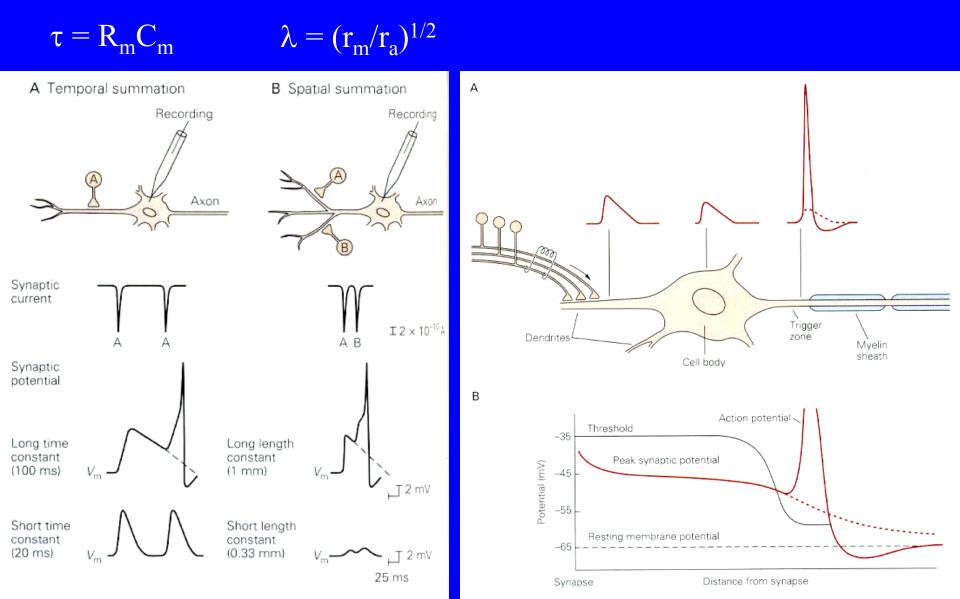
 $V_{\rm m} = \frac{RT}{F} \ln \left\{ \frac{P_{\rm K}[{\rm K}]_{\rm o} + P_{\rm Na}[{\rm Na}]_{\rm o} + P_{\rm Cl}[{\rm Cl}]_{\rm i}}{P_{\rm K}[{\rm K}]_{\rm i} + P_{\rm Na}[{\rm Na}]_{\rm i} + P_{\rm Cl}[{\rm Cl}]_{\rm o}} \right\}$

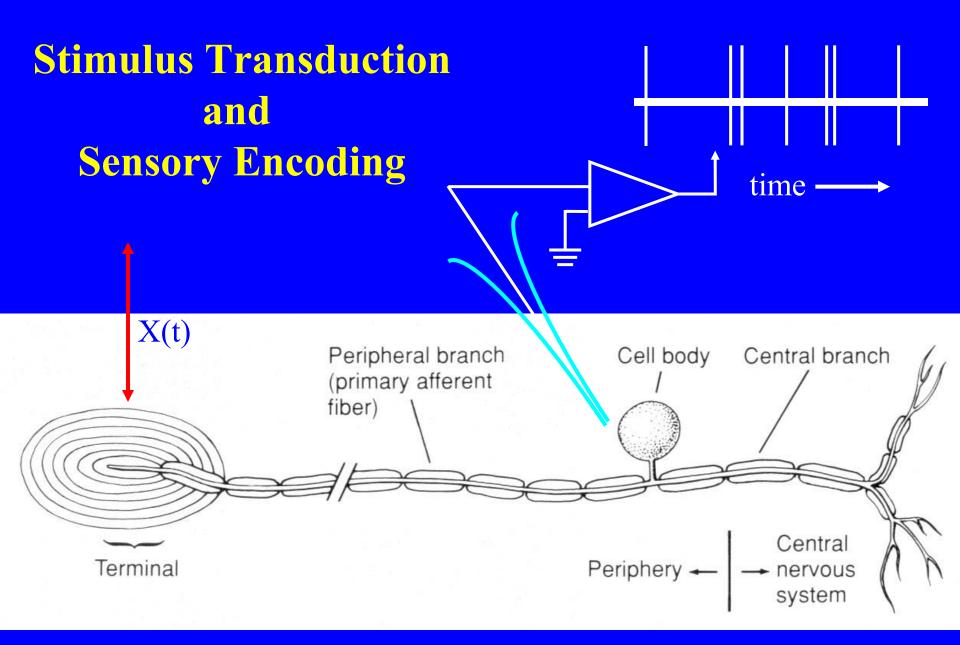
- Based on *electrochemical gradient*
- generalizes to Nernst Eq. for single species
- small effect of pump on $V_{\rm m}$, except to establish/maintain
- Na⁺/K⁺-ATPase "pump" is *electrogenic* since it moves net charge across the membrane (3Na⁺/2K⁺)
- Ion exchangers (Na⁺/Ca²⁺) use potential energy of one species to move the other species uphill

Changes in $V_{\rm m}$

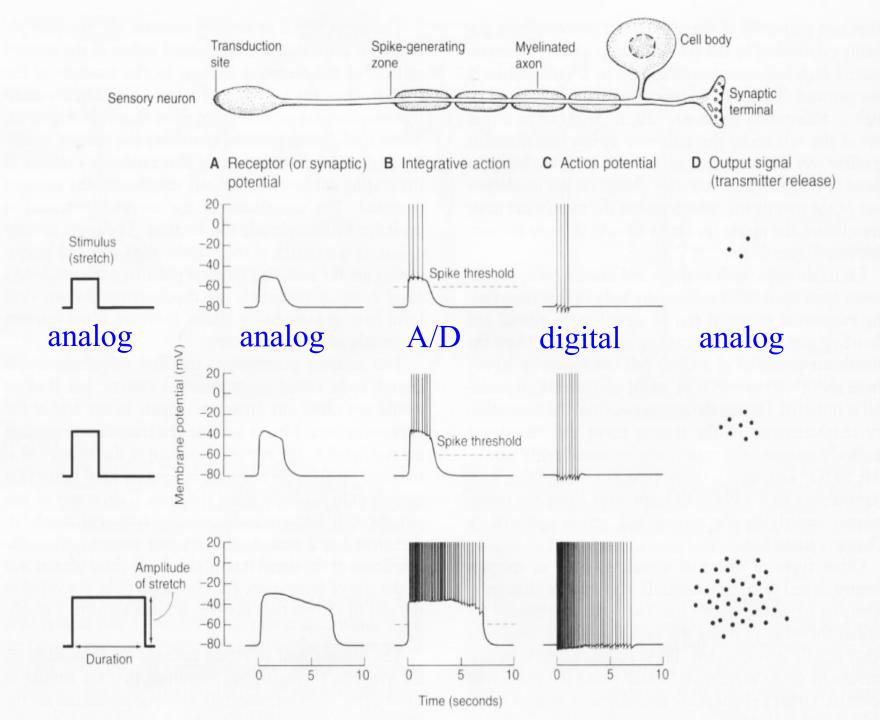
- terms:
 - *de*polarizing: $V_{\rm m}$ from rest towards 0
 - *hyper*polarizing: $V_{\rm m}$ more negative than rest
 - *re*polarizing: return to from $V_{\rm m}$ depolarization
 - overshoot: $V_{\rm m}$ positive
 - graded potential (changes) because charge moves (e.g., ions flow) across the membrane, due to changes in g. Decay is according to length constant
 - synaptic potentials
 - *generator potentials* (sensory neurons)
 - action potentials
- *Spatial Integration* of synaptic activity from different sites of the postsynaptic neurons (long vs. short space constants)
- *Temporal Integration* of same at different times (long vs. short time constants)

Length & Space Constants





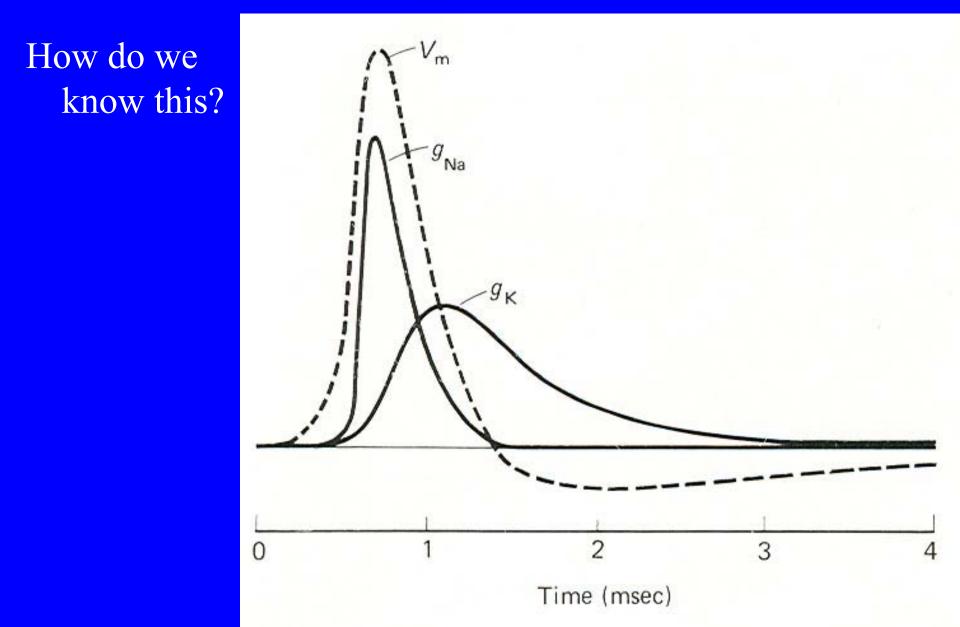
Thus, analog (continuous)-to-digital (discrete) conversion



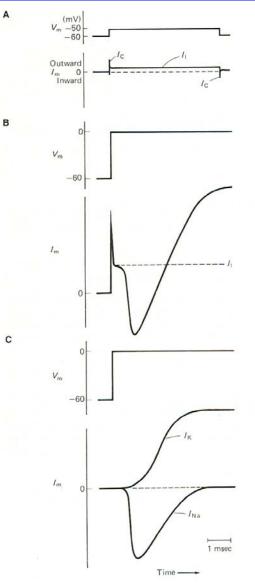
Action Potential

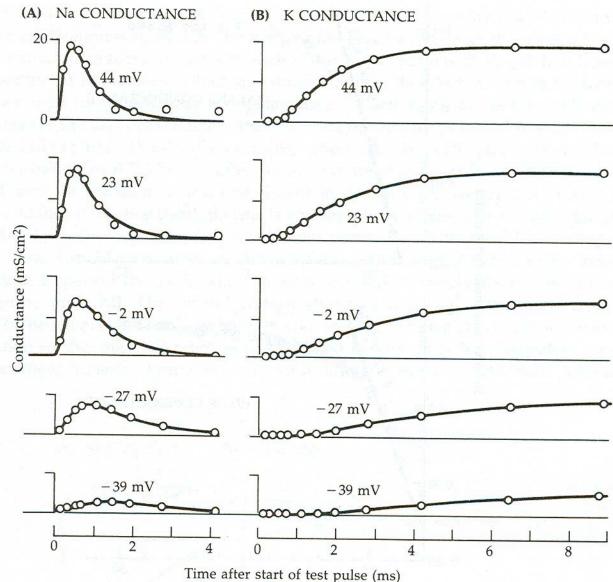
- Na: $g_{Na} = I_{Na} / (E E_{Na}); E_{Na} \sim +67 \text{ mV}$
 - opens fast, positive feedback mechanism
 - closes fast, negative feedback mechanism
 - inactivates spontaneously
- K: $g_K = I_K / (E E_K); E_K \sim -90 \text{ mV}$
 - opens slowly
 - closes v. slowly
 - inactivates from polarization only

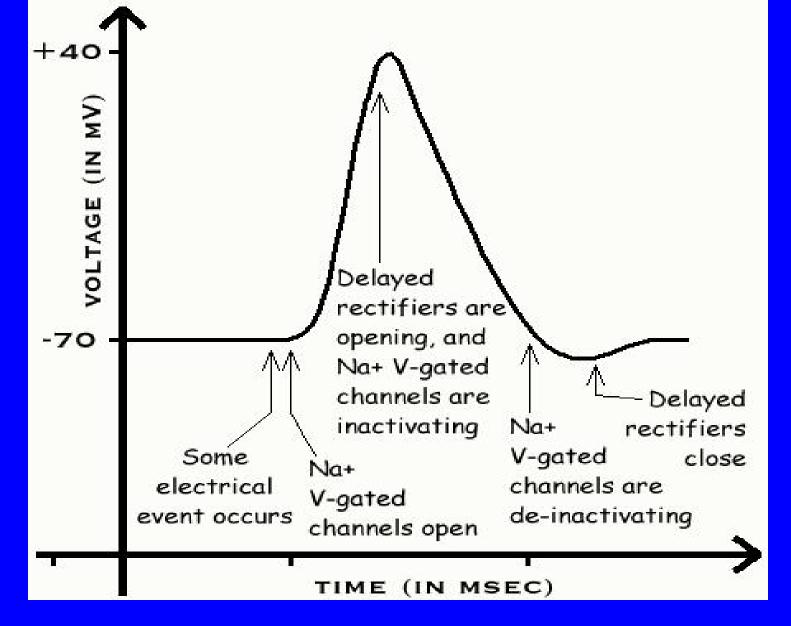
Action Potential Currents



Voltage Clamp Experiments Hodgkin & Huxley (1950s)



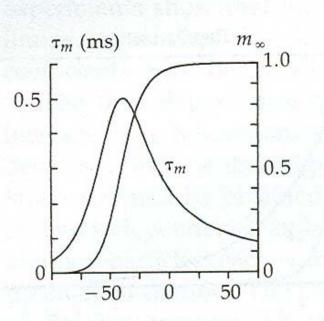


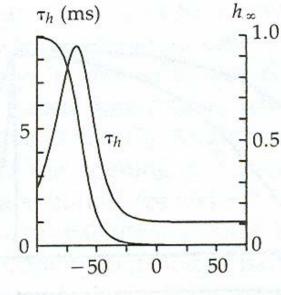


Absolute Refractory Period: due to Na⁺ wide open or inactivated Relative RP: most Na⁺ not deinactivated; K⁺ channels mostly open

Model

- Na activation and inactivation modeled as separate processes.
- K activation only.
- Basis of Na pos. feedbk



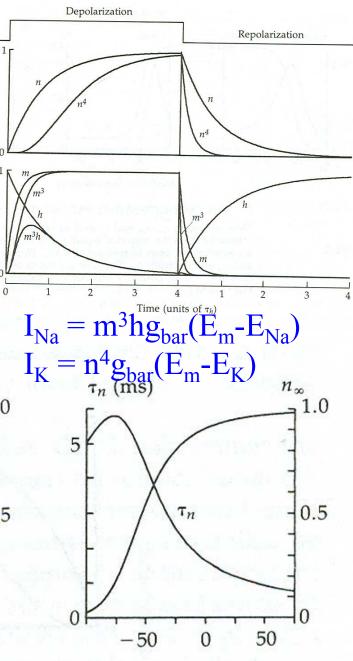


Squid axon

E_M_

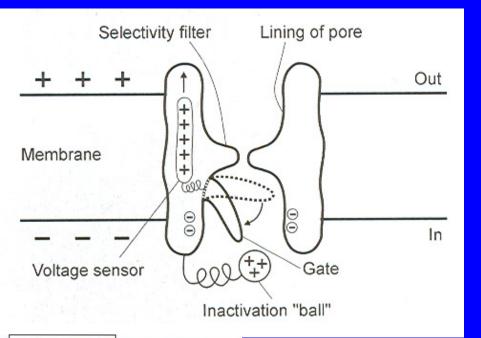
HH parameter value

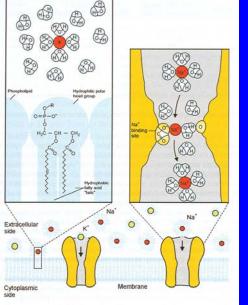
6.3°C



Membrane potential (mV)

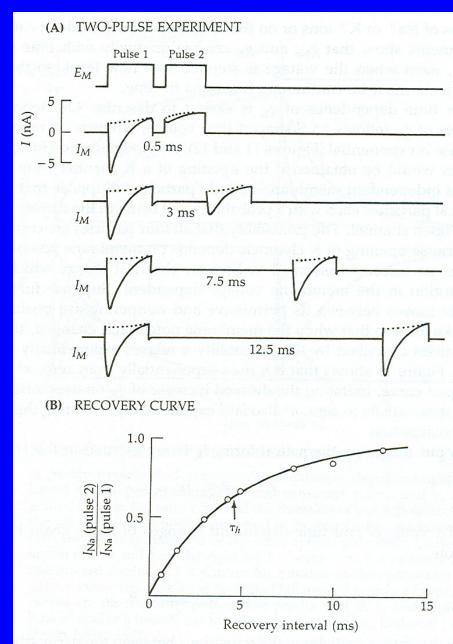
Na⁺ Channel Models



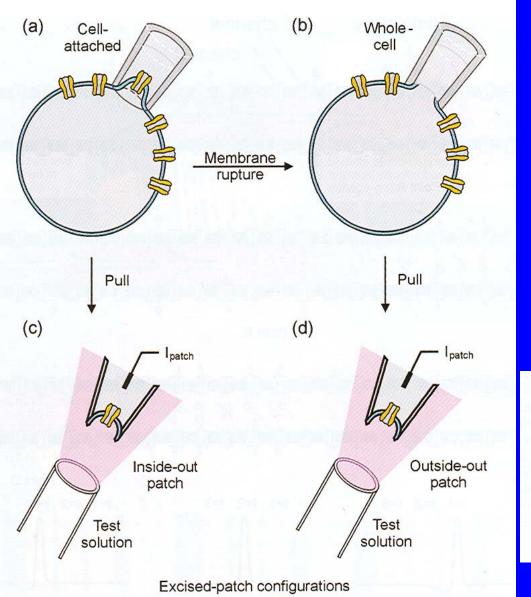


"ball & chain" inactivation
activation gate charged R groups

Inactivation

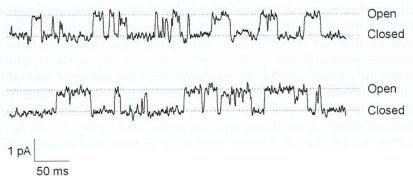


Patch Clamp Method

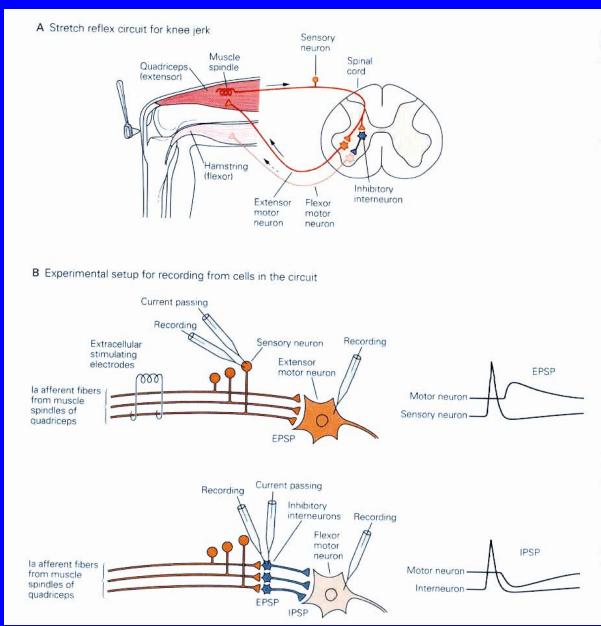


•allows for single channel recordings!

•single currents add up to whole cell currents



Synaptic Transmission

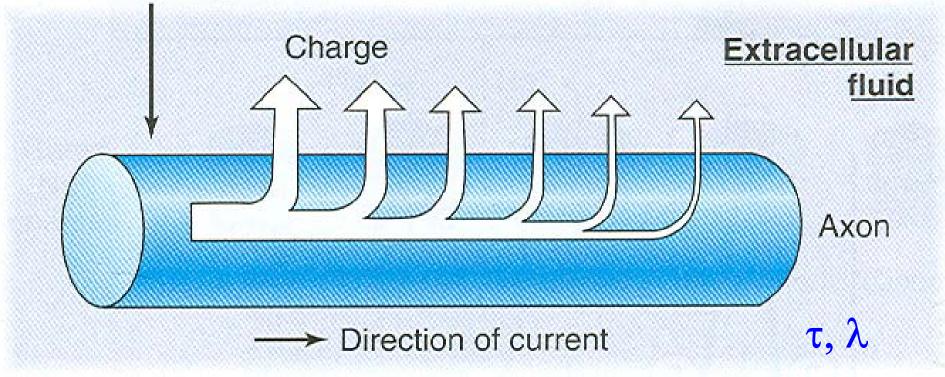


AP Propagation

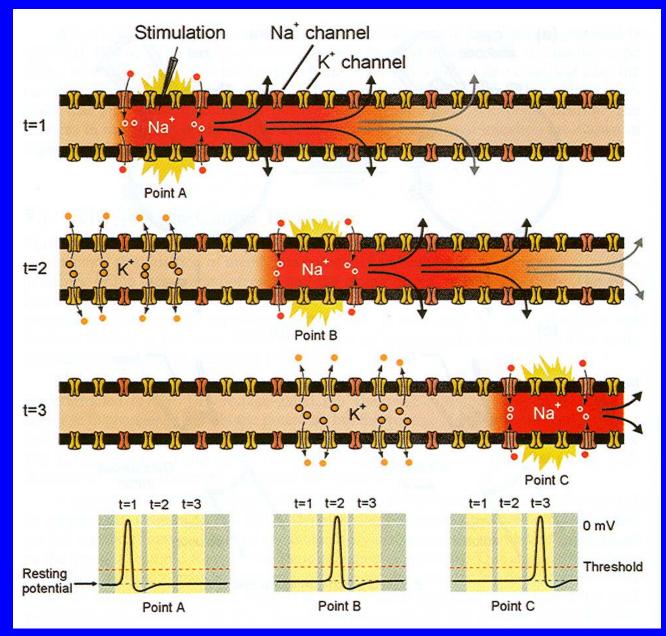
- AP depolarization acts to open neighboring Na⁺ channels, but not ones that are inactivated.
- Thus, *directional* propagation of AP occurs in axon
- APs are regenerative, not degenerative (like graded pots.).
- In myelinated axons, APs leap from one NoR to the next
 - Na⁺ chs only at NoR
 - myelin prevents leak through memb
- \uparrow mylen \rightarrow \uparrow conduction velocity
- \uparrow diam \rightarrow \uparrow conduction velocity

Passive Spread

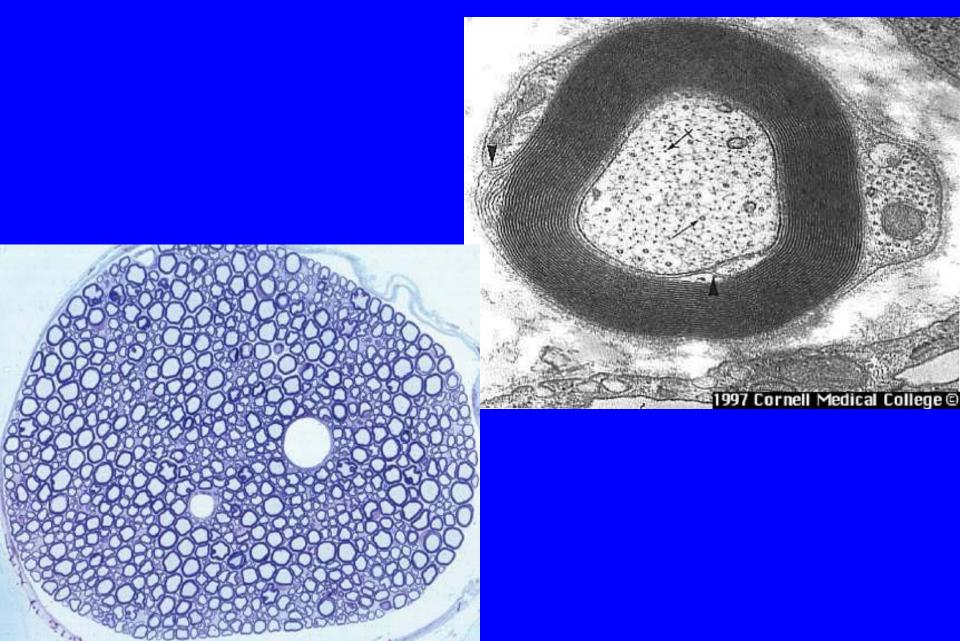
Site of initial depolarization



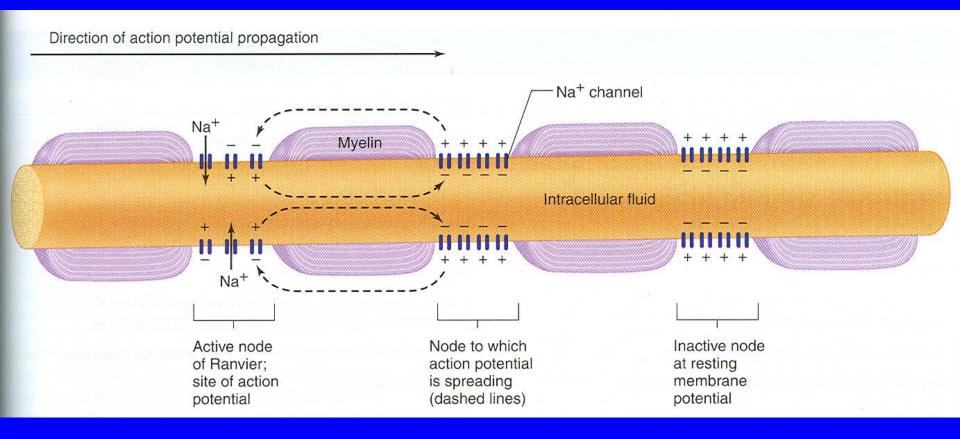
Active AP Propagation



Myelination

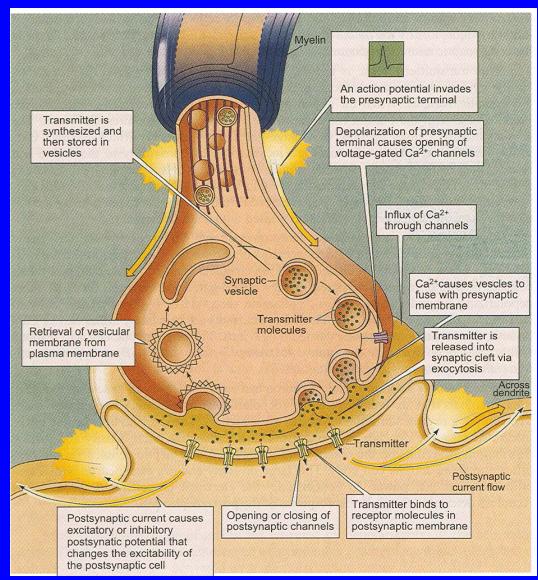


Saltatory Conduction

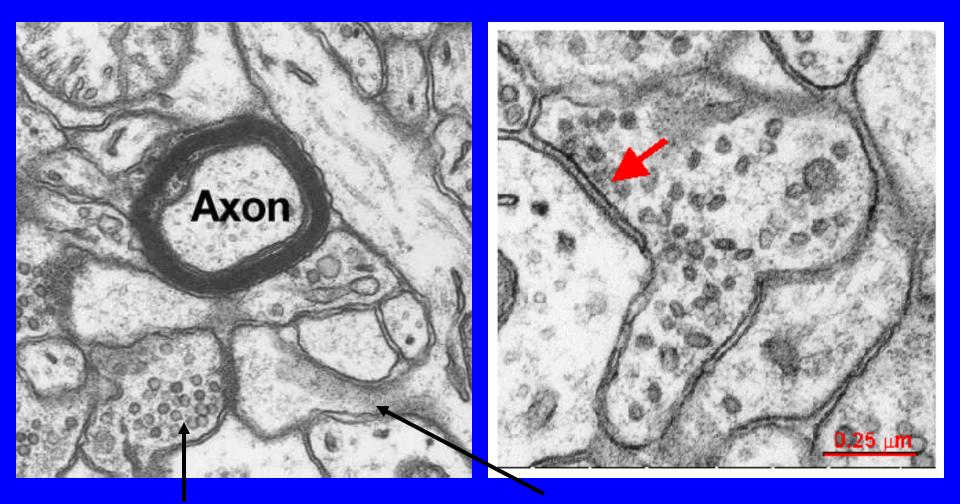


Synaptic Transmission

- 1. AP invades axon terminals
- 2. Vm-dep Ca2+ Chs open
- 3. [Ca2+]in rises
- 4. s. vesicles fuse to plasma m.
- 5. neurotransmitter released into s. cleft
- 6. NT binds to postsynaptic Rs, opening them
- 7. Cations or anions flow in *Thus, electrochemical communication*



Synapses



presynaptic vesicles

dendritic spine

What Do I Do?

- record the electrical activity of individual neurons in context, while system is functioning
- perturb systemic variables, analyze neural/network computations
- quantify information in spike trains using information theory
- record network/systemic activity using optical imaging techniques

What Can YOU Do?

- design/manufacture microrecording electrodes, arrays
- signal processing of optical imaging data
- signal processing of spike train data
- design/manufacture microchannel devices for chronic implantable devices for integration with the nervous system